

SELECTIVE α -1A ADRENERGIC RECEPTOR ANTAGONISTS. EFFECTS OF PHARMACOPHORE REGIO- AND STEREOCHEMISTRY ON POTENCY AND SELECTIVITY

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Received 8 June 1998; accepted 27 July 1998

Abstract: The anti-anxiety agent ipsapirone has been shown to have modest affinity for α -1 receptors. We disclose the discovery of potent α -1a receptor subtype selective antagonists based on the ipsapirone structure which possess selectivity versus the 5-HT receptors tested. These antagonists were obtained by tethering a saccharin ring to 4-phenyl-3-carboxyethyl piperidines. The design principles which led to this structural motif are discussed. The synthesis of key analogs, their SAR, as well as results of selected in vitro and in vivo studies are described. © 1998 Elsevier Science Ltd. All rights reserved.

Introduction

The three known families of G-protein coupled adrenergic receptors have been classified as the α -1, α -2, and β adrenoceptors. These receptors are distinguished based on sequence information, receptor pharmacology and signaling mechanisms. One of these receptor classes, the α -1, has recently been divided into three subclasses, the α -1a, α -1b, and α -1d. Their presence in animal and human tissues was confirmed when these receptors were cloned and expressed utilizing molecular biological techniques. The study of a variety of tissue preparations led to the discovery of a heterogeneous distribution of the three α -1 receptors within animal and human tissues.

Pharmacological investigations determined that nonselective α -1 antagonists were useful antihypertensive agents.³ These α -1 antagonists were also found to be therapeutically effective for the treatment of benign prostatic hyperplasia (BPH).⁴ Subsequently, it was discovered that the α -1a receptor was responsible for mediating smooth muscle contraction in the lower urinary tract.⁵ While the physiological roles of the α -1b and α -1d receptors in blood pressure or other physiological functions remain undefined in human, a selective α -1a adrenergic receptor antagonist may be a suitable candidate for the treatment of BPH since it may be devoid of cardiovascular effects associated with nonselective α -1 receptor antagonists.

Rationale

Our goal at the outset of this research was to synthesize potent and selective α -1a antagonists for the treatment of BPH. Our strategy was to convert ipsapirone, 1, an anti-anxiety agent which has modest affinity for α -1 receptors, into a potent α -1a antagonist. The approach was to replace the piperazine subunit with a variety of piperidines (Figure 1). Herein, we describe our preliminary results.

Initially, we considered replacing the N-(2-pyrimidinyl)piperazine with a 4-carboxyethyl 4-phenyl piperidine (Figure 1, A), while maintaining the butyl saccharin moiety. However, our concern about potential metabolic generation of the known opioid ligand, normeperidine, via oxidative dealkylation of the piperidine shifted our focal point to alternative 4-phenyl piperidines. Our previous findings⁶ indicated that α -1 subtype selectivity was induced by installing a carboxy group three atoms away from the biogenic amino group of a nonselective α -1 antagonist. We implemented a similar approach for dealing with α -1a antagonists containing meperidine type subunits (Figure 1, B).

Results and Discussion

Synthesis

The first series of antagonists prepared were 3-carboxyethyl-4-phenyl piperidine derivatives. In this series, the saccharin ring found in ipsapirone was replaced by 5-chlorosaccharin for improved pharmacokinetic properties.⁷ This substitution typically caused little change in α-1 receptor binding affinity. Their synthesis is highlighted in Scheme 1. A palladium mediated coupling⁸ of either phenylboronic acid or phenyltrimethyl stannane, 2, with enoltriflate 3 provided the 4-phenyl substituted material 4. The enoltriflate 3 was prepared from N-BOC-3-carboxyethylpiperidone. Subsequent double bond reduction of 4 produced the cis racemate, 5. Deprotection of 5 with HCl-EtOAc and alkylation with 7° yielded the cis racemic antagonist, 8. The enantiomers, (-)-cis 8a and (+)-cis 8b, were separated utilizing chiral HPLC (Chiralcel OD column). The (±)-trans analog of 8 was obtained from the base catalyzed epimerization of (±)-cis 5 to (±)-trans 5, followed by N-BOC deprotection with HCl-EtOAc and alkylation with 7. The racemic cis piperidines 5 were also separated into (-)-cis 5a and (+)-cis 5b and deprotected [(-)-cis 6a and (+)-cis 6b, respectively]. The piperidine, (+)-cis 6b, was alkylated with bromide 7, which produced (+)-cis 8b.

The racemic ester, (±)-cis 5, was carefully hydrolyzed to the corresponding acid (±)-cis 9, which after treatment with (S)-α-methylbenzyl amine provided a crystalline salt 10, which was suitable for X-ray diffraction (Scheme 2). The configuration of the salt 10 was determined to be the 3-(S)-carboxy-4-(S)-phenyl piperidine. The carboxylic acid 11 was generated by treating 10 with aqueous acid. The formation of the ethyl ester and N-BOC deprotection was accomplished by HCl-EtOH treatment, and the convergence with compound (-)-cis 6a was completed. Through the use of the optical rotations of key intermediates, we have determined that the (-)-cis N-BOC piperidine 5a led to the (-)-cis piperidine 6a which produced 8a, the (-)-cis enantiomer of antagonist 8.

Scheme 1.

$$\begin{array}{c} + \\ B(OH)_2 \\ or \\ SnMe_3 \end{array} \begin{array}{c} + \\ TfO \\ NBOC \end{array} \begin{array}{c} -CO_2Et \\ NBOC \end{array} \begin{array}{c} -CO_2Et \\ Pd(PPh_3)_4 \\ K_3PO_4 \\ THF/reflux \\ for ArSnMe_3 \\ Pd(OAc)_2 \\ NMP/-15-0 \ C \\ 82-93\% \end{array} \begin{array}{c} + \\ Br \\ -7 \\ O \end{array} \begin{array}{c} -CI \\ Pr_2NEt \\ DMF \\ 83\% \end{array} \begin{array}{c} -CI \\ DMF \\ 83\% \end{array} \begin{array}{c} -CI \\ DMF \\ BSW \\ -CI \\ DMF \\ BSW \\ -CI \\ -CIS \\$$

Scheme 2.

CO₂Et LiOH CO₂H (1) (S)-
$$\alpha$$
-methyl NBOC benzylamine NBOC Denzylamine (2) Crystallization 10, X-ray crystal structure $\frac{1 \text{ N HCl}}{\text{CHCl}_3}$ NBOC EtOH $\frac{\text{CO}_2}{\text{HCl}_{(g)}}$ $\frac{\text{HCl}_{(g)}}{\text{EtOH}}$ $\frac{\text{CO}_2}{\text{EtOH}}$ $\frac{\text{CO}_2}{\text{NH-HCl}}$ $\frac{\text{CO}_2}{\text{CHCl}_3}$ $\frac{\text{CO}_2$

The 4-carboxymethyl-4-phenyl piperidine analog 13 was prepared via alkylation of 12 with bromide 7 (Scheme 3).

Scheme 3.

Receptor Binding Experiments

The binding affinity to the human α -1 receptors for the synthetic compounds was measured utilizing cloned receptor binding assays (Table 1).⁵⁶ The opioid binding affinity for selected α -1a antagonists was measured utilizing ³H-DAMGO as the radioligand.¹⁰

Figure 2.

Table 1. α-1 binding data.

	$K_i(nM)$			
	α-la	α -1b	α-1d	
1	87	220	46	
8, (±)-trans	100	1400	320	
8 , (±)-cis	3.0	940	1800	
8a, (-)-cis	0.51	170	480	
8b , (+)-cis	98	>2000	1100	
13	44	2200	2000	

Structure-Activity Relationships

Replacing the N-(2-pyrimidinyl)piperazine present in ipsapirone, 1, with 4-carboxymethyl 4-phenylpiperidine, 13, led to a modest improvement in the α -1a binding affinity and enhanced α -1a receptor subtype selectivity. Relocation of the carboxyalkyl group to the 3-position of 4-phenylpiperidine caused a variety of stereochemically dependent effects. For example, the (\pm) -cis isomer of 8 has 15-fold higher affinity for the α -1a receptor than 13. However, the (\pm) -trans isomer of 8 exhibited little change in α -1 receptor binding affinity relative to 13. Assuming the piperidine present in (\pm) -trans 8 adopts a chair conformation, the 3,4-substituents should reside in a diequatorial arrangement. If the conformation of the piperidine ring present in (\pm) -cis 8 is consistent with that of

10 (determined by X-ray diffraction), the 4-phenyl group in (\pm) -cis 8 would occupy an axial position and the 3-carboxylate an equatorial post (Figure 2). In this conformation, the more potent cis isomer of 8 may benefit from either a unique hydrogen bond between the equatorial 3-carboxylate and the α -1a receptor or a better fit for the axial 4-phenyl group within the α -1a receptor.

The (-)-cis enantiomer 8a has subnanomolar affinity for the α -1a receptor and is highly selective against the α -1b and α -1d receptors, while 8b has much lower affinity for the α -1a receptor. Assuming that the two enantiomers of (\pm)-cis 8 bind in a similar manner, (-)-cis 8a may benefit from a favorable interaction between the 3-carboxylate and the α -1a receptor and/or (+)-cis 8b may suffer from steric or electrostatic interference with certain elements within the α -1a receptor binding site. Nevertheless, (-)-cis 8a represents the preferred α -1a antagonist within this series based on the α -1 receptor binding profile and when counterscreened, was inactive (>30 μ M) in the opioid binding assay and >500-fold selective against the other G-protein-coupled receptors (human α -2, β -1, 2, and 3, dopamine-2 and -5, and 5-HT receptors) tested. The study of the functional activity of (-)-cis 8a in isolated rat prostate tissue revealed that (-)-cis 8a competitively antagonizes phenylephrine induced contraction with a K, value of 28 nM.

We also measured the opioid binding for the putative piperidine metabolites of each antagonist. These results are summarized in Table 2.

Table 2. Opioid binding data for piperidines.

		<u>K_i (nM)</u> ³ H - DAMGO
6, (±)-tran	° CO₂Et	24,000
6a , (-)-cis	CO ₂ Et	7,600
6b , (+)-cis	CO ₂ Et	12,000
12	CO₂Et NH	1,300

The relocation of the carboxyethyl group present in 4-carboxyethyl 4-phenylpiperidine, 12 to the piperidine 3-position caused a 6- to 20-fold decrease in opioid receptor binding affinity. This decrease in opioid binding activity was stereochemically dependent. The piperidine, 6a, present in the most preferred α -la antagonist 8a, was approximately 6-fold less avidly bound than normeperidine, 12.

In Vivo Study

Prior to the delineation of the stereochemistry of the preferred enantiomer of 8, we studied the bioavailability of (\pm) -cis 8 in rats. Although reasonable plasma levels of 8 were detected its pharmacokinetic profile was poor, with 9.2% bioavailability and an 80 minute half-life.

Conclusion

The α -1a potency and selectivity of ipsapirone was increased by replacing 1-N-(2-pyrimidinyl)piperazine with the (-)-cis 3-carboxyethyl-4-aryl piperidine, (-)-cis 6a, while the 5-HT activity was diminished. The (\pm)-cis 3-carboxyethyl-4-phenyl piperidine derivative, (\pm)-cis 8, was substantially more potent and selective than the corresponding (\pm)-trans isomer, (\pm)-trans 8. The separation of the (\pm)-cis enantiomers of 8 provided a more potent and selective isomer, (-)-cis 8a, than the racemate, and a less active enantiomer, (+)-cis 8b. Therefore, the stereospecific installation of the carboxylate group at the piperidine 3-position in a relative orientation cis to the 4-phenyl substituent plays an important role in optimizing α -1a binding affinity and subtype selectivity within this class of α -1 antagonists.

The preparation of **8a** represents yet another example of the approach to design new, potent, and selective G-protein coupled receptor antagonists from existing, low-affinity, nonselective receptor ligands.

Acknowledgment

The authors thank Patrick Pollard and Meng-Hsin Chen for providing samples of 6 and 11, Sean P. McKee for preparing 1, and Richard G. Ball for solving the X-ray crystal structure of 10.

References

- Hieble, J. P.; Bylund, D. B.; Clarke, D. E.; Eikenburg, D. C.; Langer, S. Z.; Lefkowitz, R. J.; Minneman, K. P.; Ruffolo, R. R. Pharmacol. Rev. 1995, 47, 267.
- (a) Schwinn, D. A.; Lomasney, J. W.; Lorenz, W.; Szklut, P. J.; Fremeau, R. T.; Tany-Feng, T. L.; Caron, M. G.; Lefkowitz, R. J.; Cotecchia, S. J. Biol. Chem. 1990, 265, 8183.
 (b) Schwinn, D. A.; Johnston, G. I.; Page, S. O.; Mosely, M. J.; Wilson, K. H.; Worman, N. P.; Campbell, S.; Fidock, M. D.; Furness, L. M.; Parry-Smith, D. J.; Peter, B.; Bailey, D. S. J. Pharmacol. Exp. Ther. 1990, 265, 8183.
- 3. For example, Prazosin: Graham, R. M; Pettinger, W. A. N. Engl. J. Med. 1979, 300, 232.
- (a) For phenoxybenzylamine: Caine, M.; Pfau, A.; Perlberg, S. Br. J. Urol. 1976, 48, 255. (b) for terazosin: Debruyne, F. M. J.; Witjes, W. P. J.; Fitzpatrick, J.; Kirby, R.; Kirk, D.; Prezioso, D. Eur. Urol. 1996, 30, 369. (c) for prazosin: Kirby, R. S.; Coppinger, S. W. C.; Gorcoran, M. O.; Chapple, C. R.; Flannagan, M.; Milroy, E. T. G. Brit. J. Urol. 1987, 60, 136.
- (a) Price, D. T.; Schwinn, D. A.; Lomasney, J. W.; Allen, L. F.; Caron, M. G.; Lefkowitz, R. J. J. Urol. 1993, 150, 546.
 (b) Forray, C.; Bard, J. A.; Wetzel, J. M.; Chiu, G.; Shapiro, E.; Tang, R.; Lepor, H.; Hartig, P. R.; Weinschank, R. L.; Branchek, T. A.; Gluchowski, C. Mol. Pharmacol. 1994, 45, 703.
 (c) Chapple, C. R.; Burt, R. P.; Andersson, P. O.; Greengrass, P.; Wyllie, M.; Marshall, I. Br. J. Urol. 1994, 74, 585.
- Patane, M. A.; Scott, A. L.; Broten, T. P.; Chang, R. S. L.; Ransom, R. W.; DiSalvo, J.; Forray, C.; Bock, M. G. J. Med. Chem. 1998, 41, 1205.
- 7. Erb, J. M.; Lee, H.Y.; Munson, P. M.; Nerenberg, J. B.; Thompson, W. J. unpublished results.
- 8. Fu, J.M.; Sniekus, V. Tetrahedron Lett. 1990, 31, 1665.
- 9. Desai, R. C.; Hlasta, D. J.; Monsour, G.; Saindane, M. T. J. Org. Chem. 1994, 59, 7161.
- Slater, P.; Cross, A. J. In Neuropeptide Technology. Gene Expressions and Neuropeptide Receptors; Conn, P. M., Ed.; Academic: San Diego, 1991; Vol. 5 pp 459-478.